



# Transducer Arrays for Ultrasonic Particle Manipulation

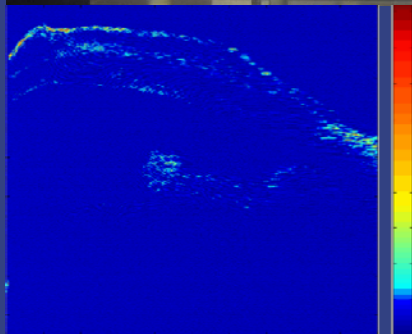
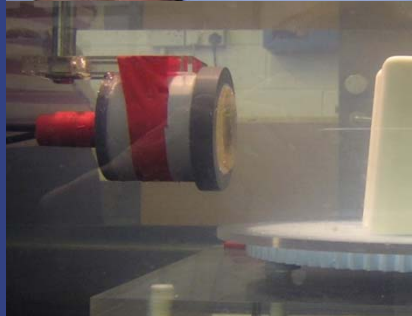
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School of Engineering Sciences  
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# Outline

Introduction

Ultrasonic particle manipulation

Finite element modelling

Experimental validation

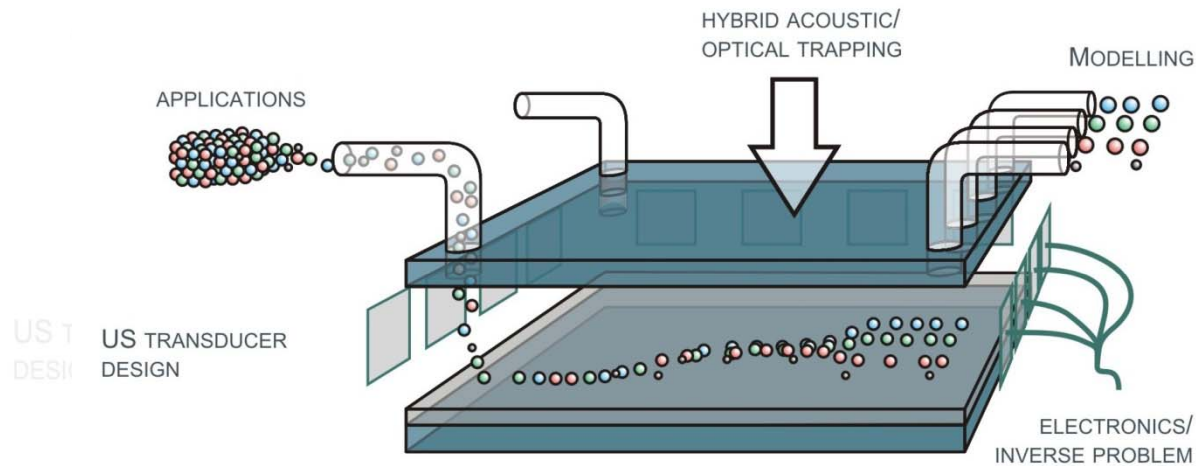
Conclusions



# INTRODUCTION



## Electronically controlled acoustic particle manipulation for life sciences research



HIGH FREQUENCY US  
TRANSDUCER FABRICATION





# Electronic Sonotweezers

- Ultrasonic particle manipulation tools enable:
  - Manipulation of **larger particles**, **cells** and **groups of cells** compared to other manipulation technologies
  - Dimensions of less than 1  $\mu\text{m}$  up to hundreds of microns
- Ultrasound devices are readily **integrated** with **microfluidic devices** and **electronics**
- Aim of Sonotweezers:

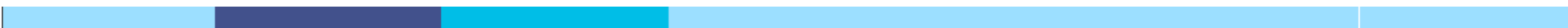
**Dexterous acoustic manipulation**





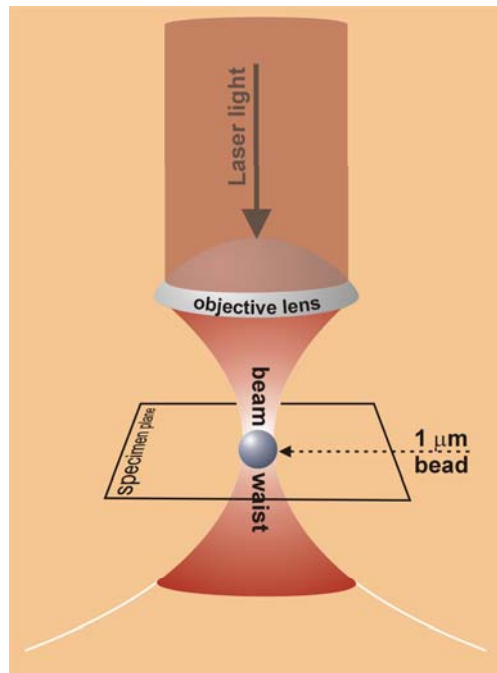
# Electronic Sonotweezers

- Applications of **Sonotweezers** include:
  - Separation and sorting of cells
  - Investigation of cell characteristics
  - Measurements of cell forces
  - Tissue engineering
  - Positioning cells at sensors
- Many applications require manipulation force in more than one direction



# Current Tweezing Technology

## Optical Tweezing

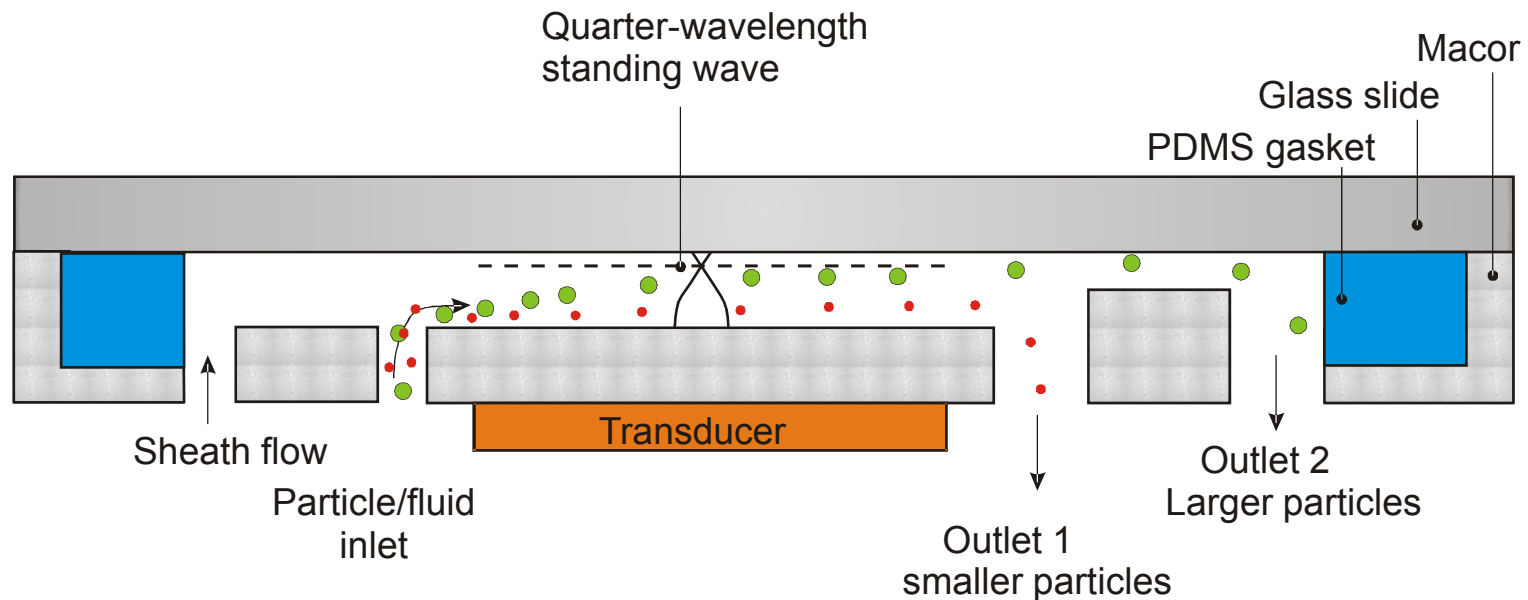


02-02-03  
OT Levovist CA  
LG l=2 Hologram  
x100 obs. obj.  
x63 tweezing obj.

- Lateral manipulation possible by steering optical trap with mirrors

# Ultrasonic Particle Manipulation

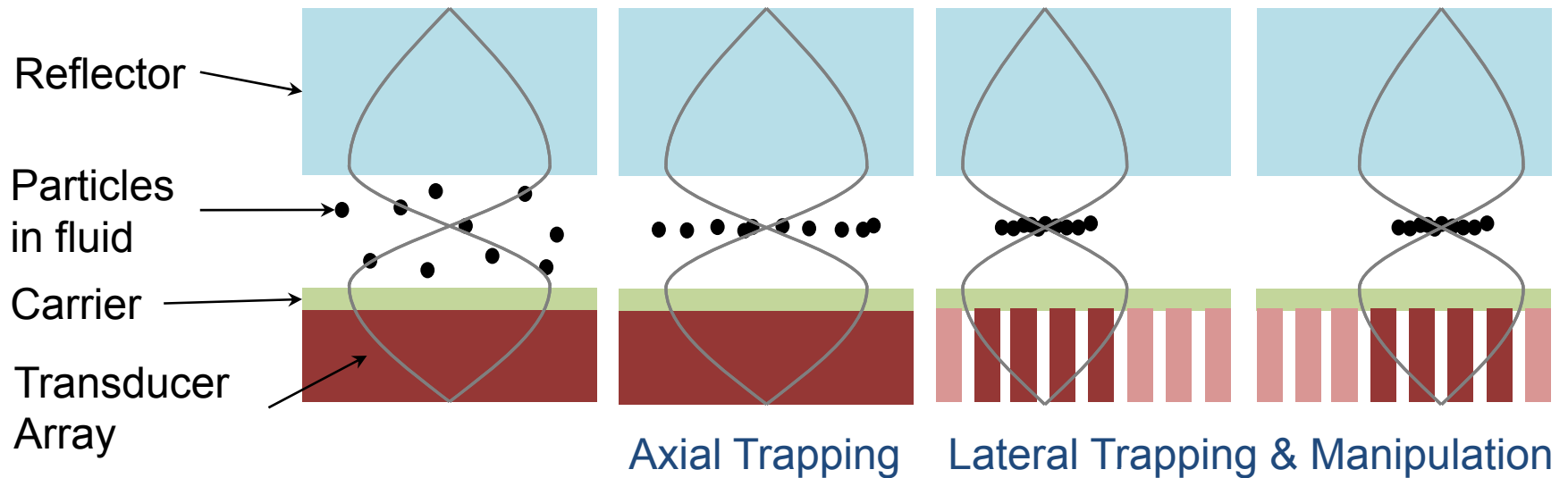
- Transducer forms ultrasound standing wave (USW) in channel
- Acoustic field is effectively constant along length of channel



P. Glynne-Jones et. al., "Ultrasonic radiation forces for cell sorting and characterisation," UIA Symposium, Glasgow, UK, 23 May 2011.



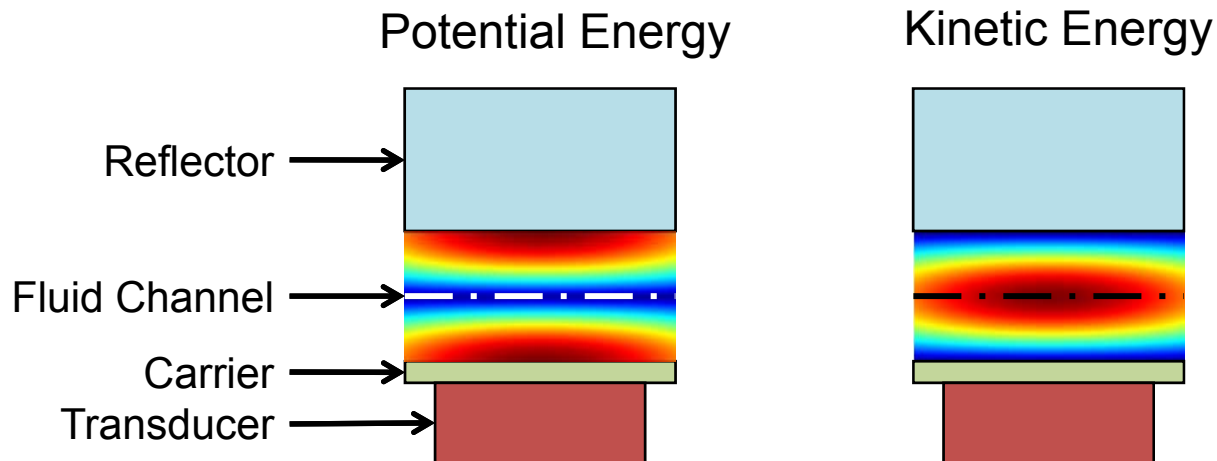
# Ultrasonic Manipulation with Arrays



- Array replaces single transducer in USW device
- More dexterity than USW device with single element
- Can manipulate larger particles than optical tweezers

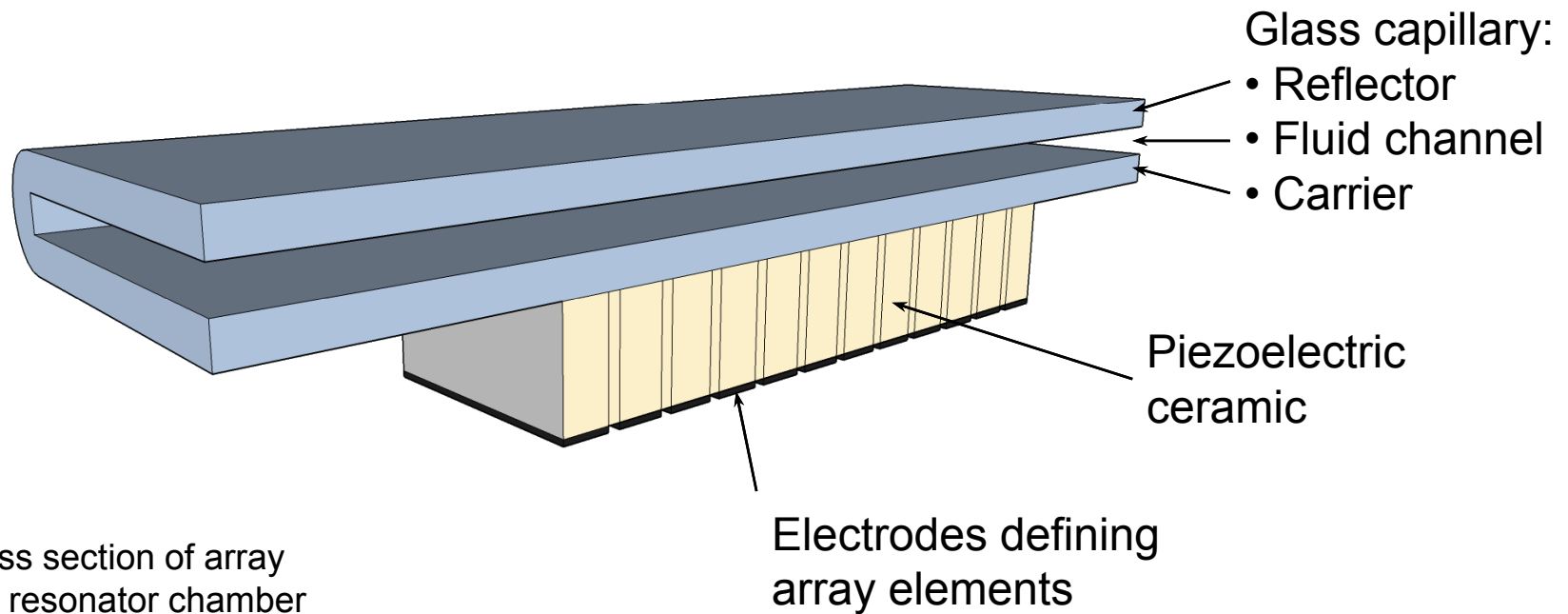
# Ultrasonic Manipulation Forces

- Force on particles is towards:  
Pressure node  $\leftrightarrow$  Potential energy minimum  
Velocity maximum  $\leftrightarrow$  Kinetic energy maximum



$$F = -\nabla(A \times E_{pot} - B \times E_{kin})$$

# Resonator and Array Design



Resonance frequency: 2.5 MHz

Channel thickness: 300  $\mu\text{m}$

Reflector thickness: 300  $\mu\text{m}$

Transducer thickness: 1 mm

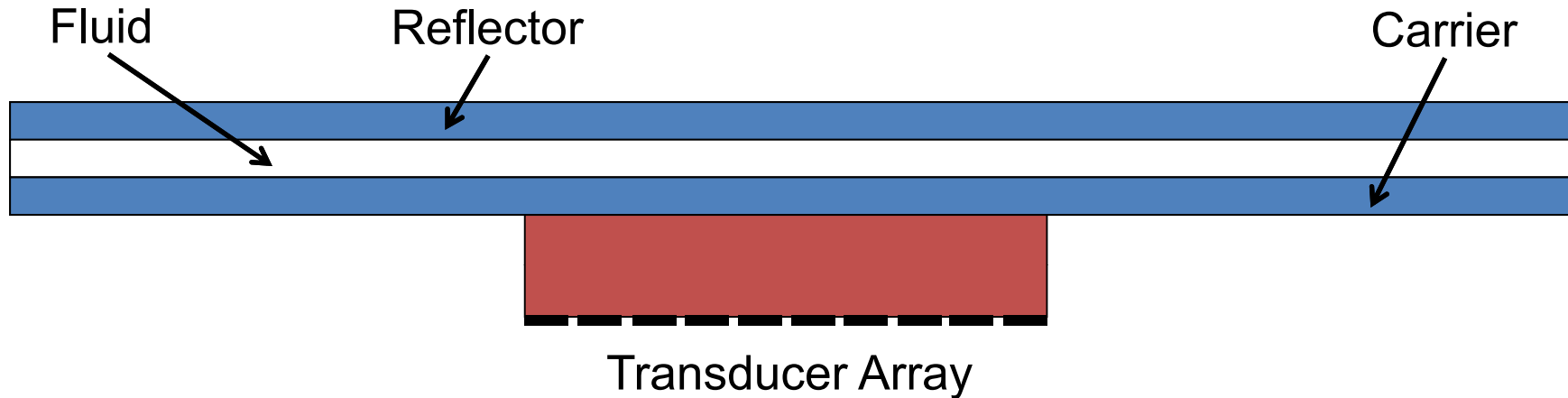
Element pitch: 500  $\mu\text{m}$



# FINITE ELEMENT MODELLING



# Finite Element Model

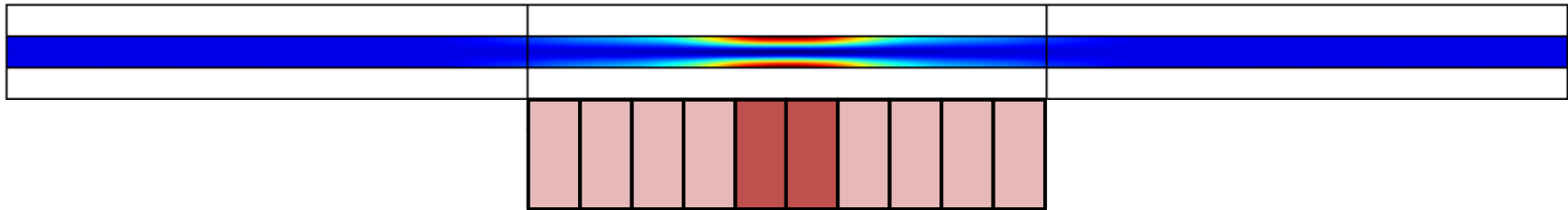


Layer	Material	Thickness
Reflector	Glass	300 $\mu\text{m}$
Fluid	Water	300 $\mu\text{m}$
Carrier	Glass	300 $\mu\text{m}$
Transducer	PZ26	1000 $\mu\text{m}$

# Acoustic Energy Distributions

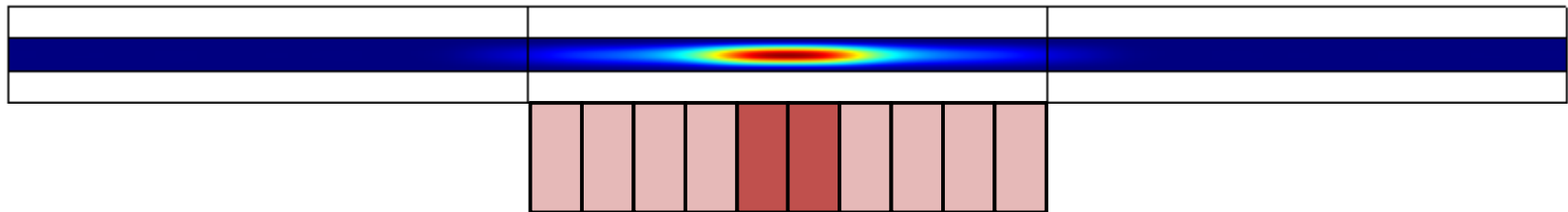
Potential Energy

Trapping position: at minimum



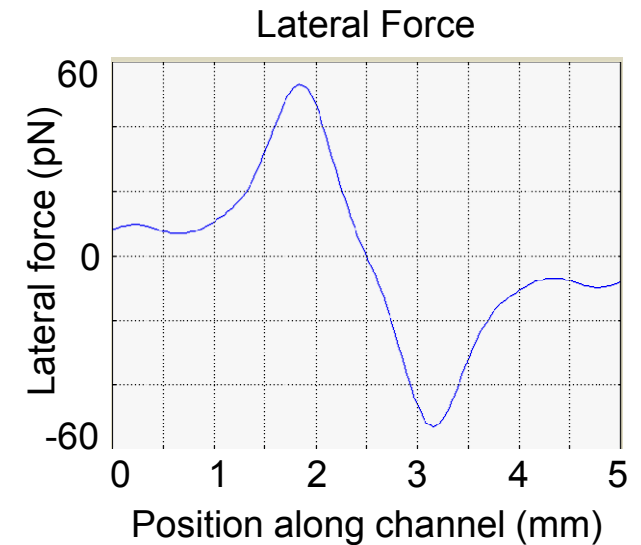
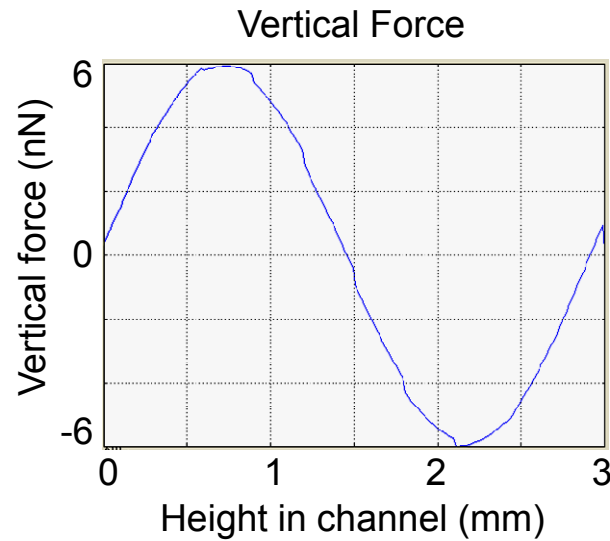
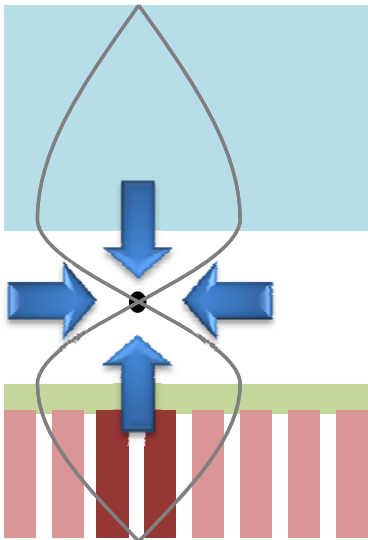
Kinetic Energy

Trapping position: at maximum



Energy Density: Min  Max

# Calculated Force Distributions



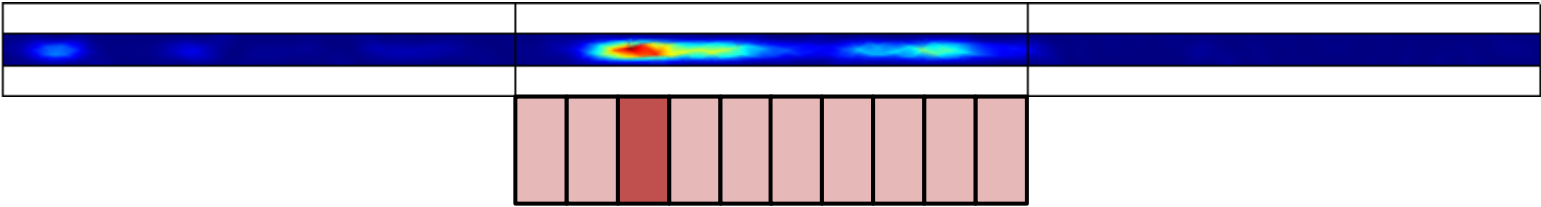
$$F = -\nabla (A \times E_{pot} - B \times E_{kin})$$

The equation is annotated with blue circles around the terms  $A \times E_{pot}$  and  $B \times E_{kin}$ . Two blue arrows point from these circles towards the top-left and top-right corners of the slide, respectively.

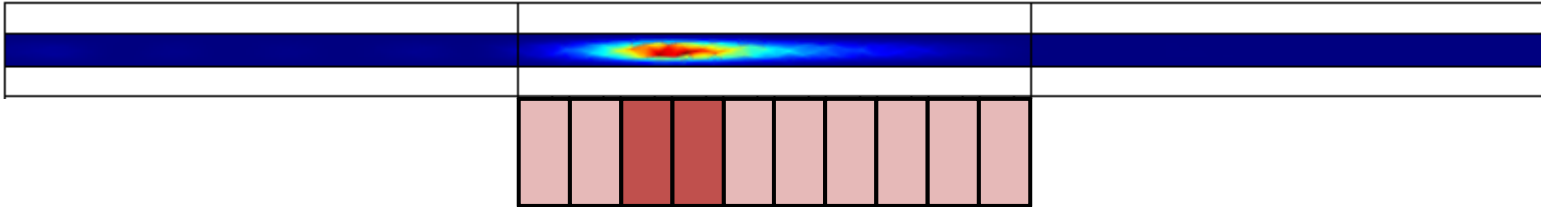
Vertical / Lateral Force: **100**

# Kinetic Energy Densities

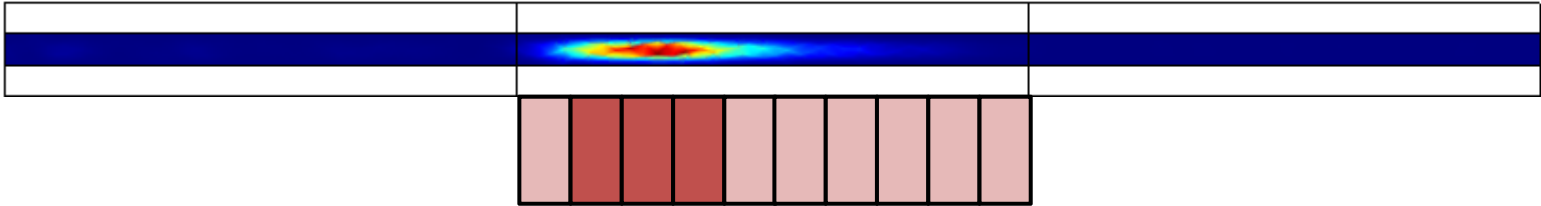
1 Element Active



2 Elements Active



3 Elements Active



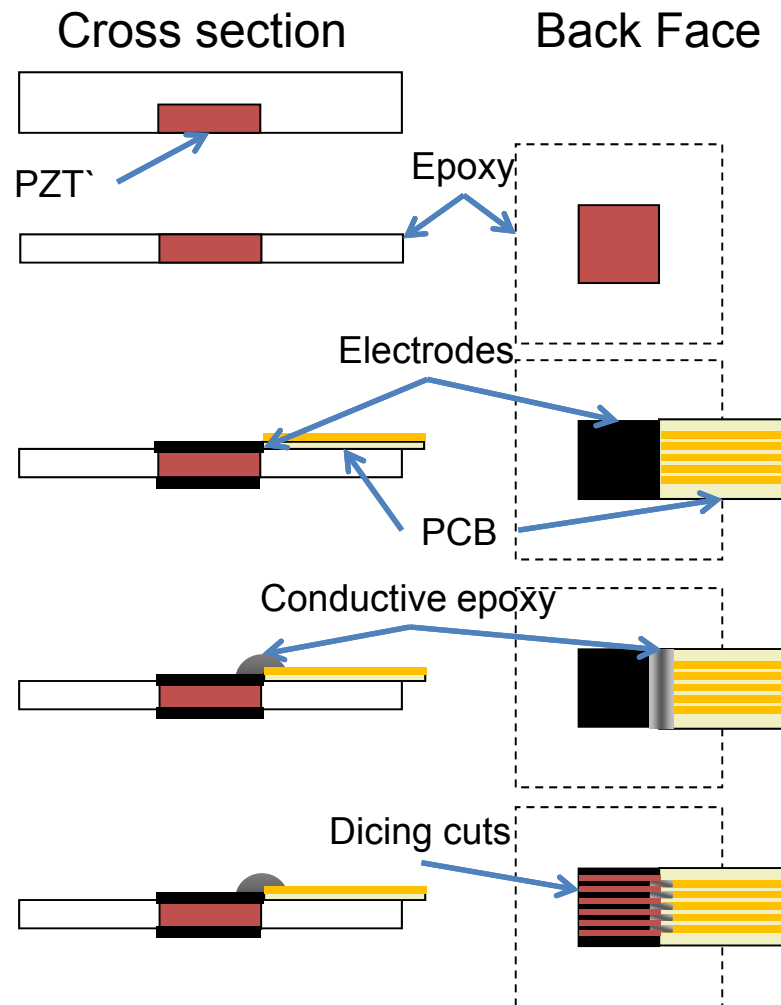




# **EXPERIMENTAL VALIDATION**



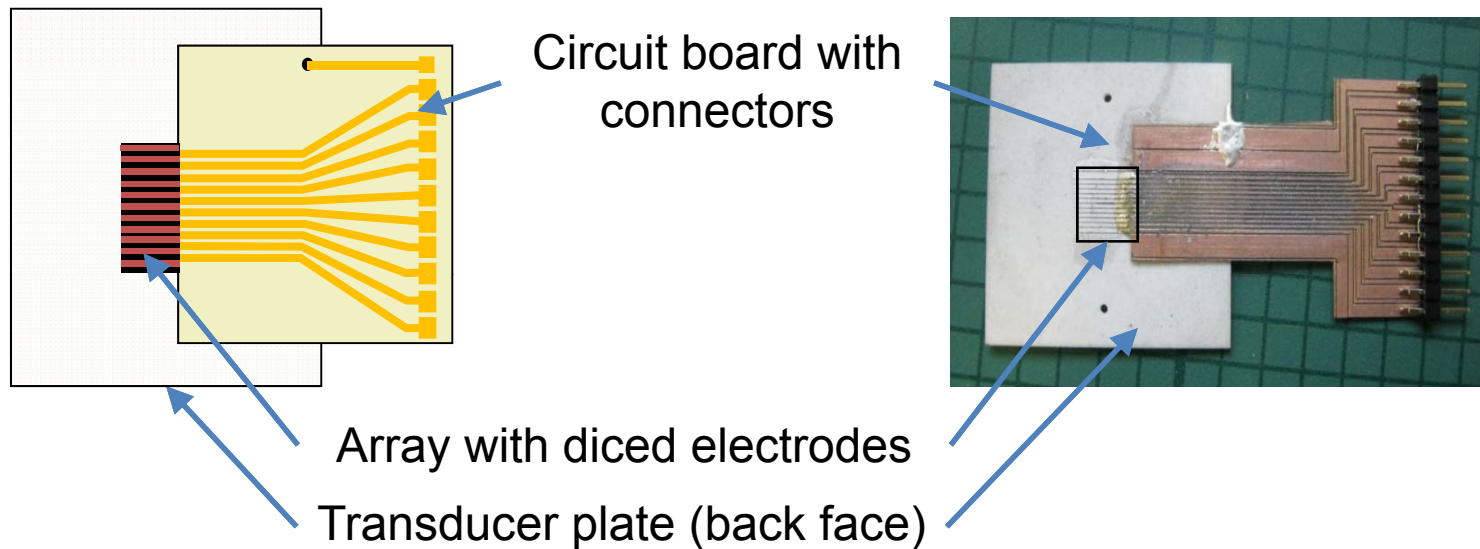
# 1D Array Fabrication



## Fabrication Process

1. Embed piezoceramic plate in microballoon loaded epoxy
2. Lap transducer plate
3. Deposit electrodes on surfaces of piezoceramic plate
4. Affix PCB to back face of transducer substrate
5. Connect tracks on PCB to transducer electrode with conductive epoxy
6. Dice through transducer electrode and conductive paste to separate element electrodes

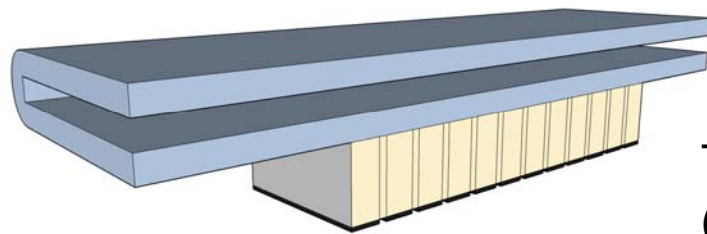
# Fabricated Array



Transducer dimensions	4 mm x 6 mm
Transducer plate thickness:	1 mm
Element pitch:	500 $\mu\text{m}$
Array elements:	12

# Experimental Setup

- Capillary coupled to array with glycerol
- Capillary channel filled with suspension of 10  $\mu\text{m}$  fluorescent polystyrene beads in water
- Drive: 2 elements
  - 17 Volts CW @ 2.408 MHz
  - Connection switched along array

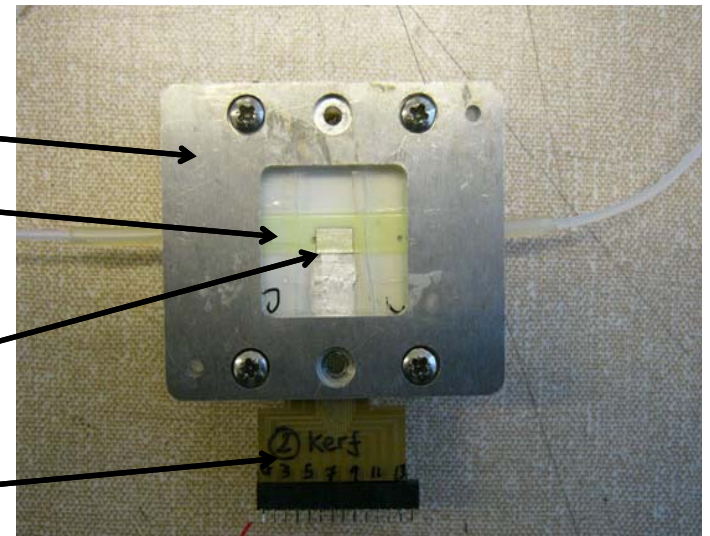


Housing

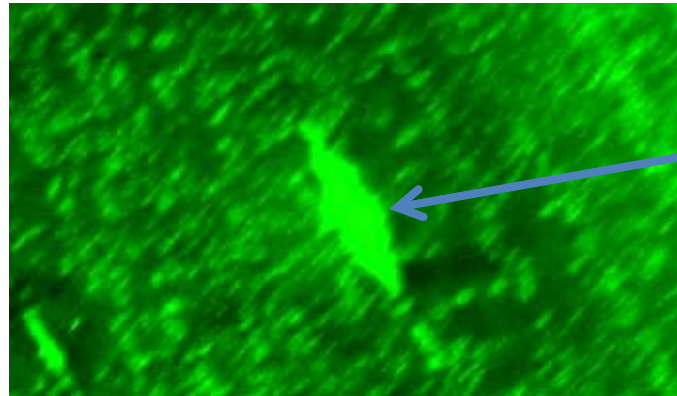
Glass capillary

Transducer Plate  
(front face)

PCB and  
connectors



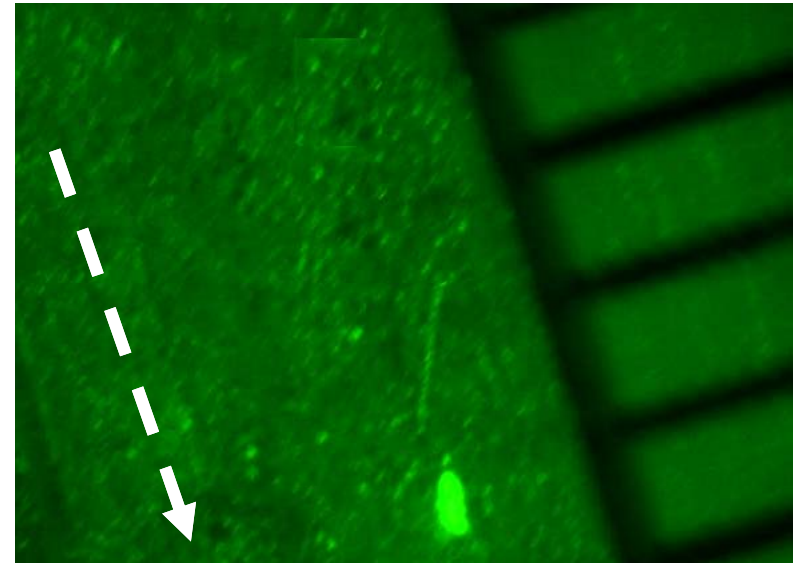
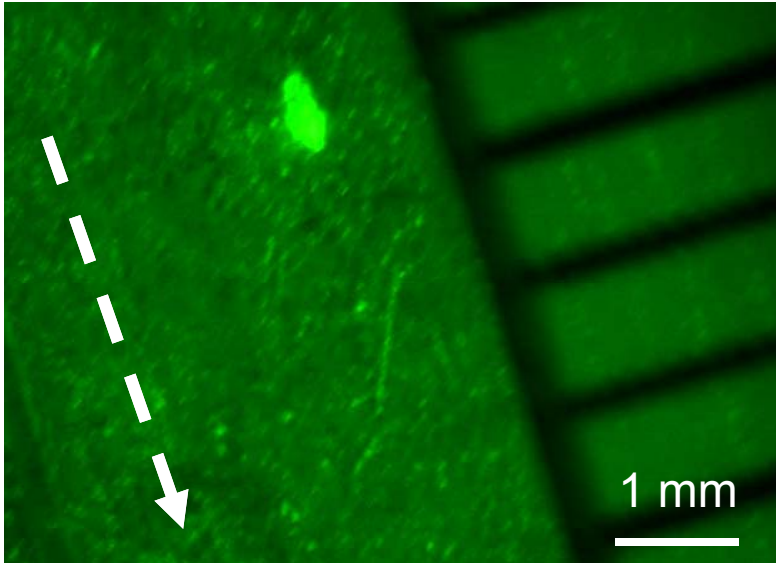
# Results: Trapping Particles



Agglomerate of  
10  $\mu\text{m}$  beads

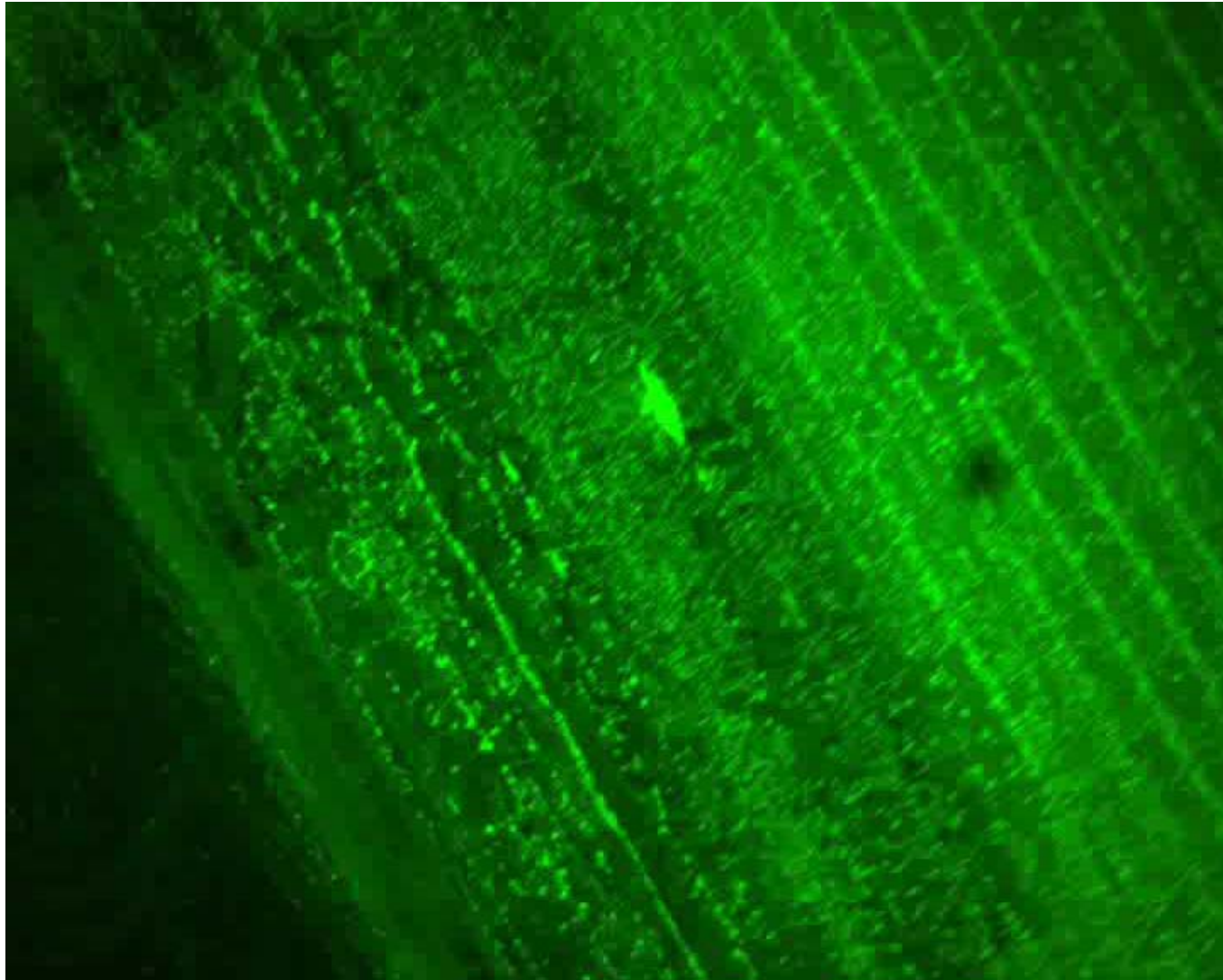
Agglomerate length:	650 $\mu\text{m}$
Vertical force measurement:	180 pN
• Balance acoustic radiation force with gravity	
Lateral force measurement:	2 pN
• Determine force from drag on particle	
Vertical / Lateral force:	<b>90</b>

# Results: Moving of Beads in Channel



- Connector switched along 6 elements
- Beads move 3 mm along channel
- Particles moved smoothly and consistently along microfluidic channel

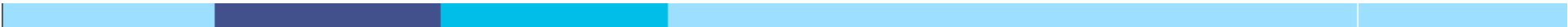
# Results: Moving beads along channel





# Summary & Conclusions

- 2.5 MHz, **1-D array** in planar resonator developed for acoustic particle or cell manipulation in microfluidic device.
  - Resonator structure forces particles to centre of fluid channel
  - Switching subset of active elements forces particles to centre of active area and moves particles along channel
- Simulation confirms expected trapping points at **pressure node** and **velocity maximum**
- Experiment demonstrates feasibility of **electronically controlled lateral manipulation**







# Industrial Collaborators

- Agilent Technologies
  - Genetix Ltd.
  - Loadpoint Ltd.
  - Logitech Ltd.
  - PCT Ltd.
  - UK Defence Science and Technology Labs
  - Weidlinger Associates, Inc.
- 



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